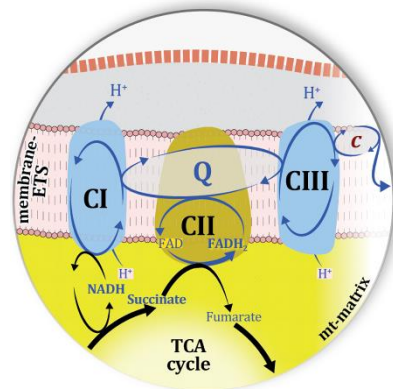


Theoretical Communication

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Complex II ambiguities – FADH₂ in the electron transfer system

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Summary

The prevailing notion that reduced cofactors NADH and FADH₂ transfer electrons from the tricarboxylic acid cycle to the mitochondrial electron transfer system creates ambiguities regarding respiratory Complex II (CII). The succinate dehydrogenase subunit SDHA of CII oxidizes succinate and reduces the covalently bound prosthetic group FAD to FADH₂ in the canonical forward tricarboxylic acid cycle. However, several graphical representations of the electron transfer system depict FADH₂ in the mitochondrial matrix as a substrate to be oxidized by CII. This leads to the false conclusion that FADH₂ from the β-oxidation cycle in fatty acid oxidation feeds electrons into CII. In reality, dehydrogenases of fatty acid oxidation channel electrons to the coenzyme Q-junction but not through CII. The ambiguities surrounding Complex II in the literature and educational resources call for quality control, to secure scientific standards in current communications of bioenergetics, and ultimately support adequate clinical applications. This review aims to raise awareness of the inherent ambiguity crisis, complementing efforts to address the well-acknowledged issues of credibility and reproducibility.

1. Introduction

Current studies on cellular and mitochondrial bioenergetics sparked a new interest in the tricarboxylic acid (TCA) cycle – the citric acid cycle or Krebs cycle (Krebs, Eggleston 1940; Gnaiger et al 2020; Bénil et al 2022; Arnold, Finley 2023). TCA cycle metabolites are oxidized while reducing NAD⁺ to NADH+H⁺ in the forward cycle, or are transported into the cytosol (Murphy, O'Neill 2018). Respiratory Complex II (CII, succinate dehydrogenase SDH; succinate-ubiquinone oxidoreductase; EC 1.3.5.1) has a unique position in both the TCA cycle and the mitochondrial membrane-bound electron transfer system (membrane-ETS). All genes for CII are nuclear encoded, with exceptions in red

algae and land plants (Huang et al 2019; Moosavi et al 2019). Succinate:quinone oxidoreductases (SQRs, succinate dehydrogenases SDH) favour oxidation of succinate and reduction of quinone in the canonical forward direction of the TCA cycle and electron transfer into the Q-junction (Cecchini 2003). Operating in the reverse direction, quinol:fumarate reductases (QFRs, fumarate reductases, FRD) reduce fumarate and oxidize quinol (Iverson 2013; Maklashina et al 2022). The reversed TCA cycle has gained interest in studies ranging from metabolism in anaerobic animals (Hochachka, Somero 2002; Müller et al 2012), thermodynamic efficiency of anaerobic and aerobic ATP production (Gnaiger 1993), reversed electron transfer and production of reactive oxygen species (Tretter et al 2016; Robb et al 2018; Spinelli et al 2021), hypoxia and ischemia-reperfusion injury (Couchani et al 2014), to evolution of metabolic pathways (Lane 2022). In cancer tissue CII plays a key role in metabolic remodeling (DeBerardinis, Chandel 2016; Schöpf et al 2020). Beyond its role in electron transfer in the TCA cycle and the membrane-ETS, CII and succinate serve multiple functions in metabolic signaling (Murphy, Chouchani 2022; Iverson et al 2023).

The pyridine derivative NAD^+ is reduced to $\text{NADH}+\text{H}^+$ during the oxidation of pyruvate and through redox reactions catalyzed by TCA cycle dehydrogenases (DH) including isocitrate DH, oxoglutarate (α -ketoglutarate) DH, and malate DH. In turn, $\text{NADH}+\text{H}^+$ are the substrates for the oxidation reaction catalyzed by CI which is linked to reduction of the prosthetic group FMN to FMNH_2 and regeneration of NAD^+ . Likewise, the prosthetic group FAD is reduced to FADH_2 during oxidation of succinate by CII (succinate DH). Confusion emerges, however, when NADH and FADH_2 are considered as the reduced substrates feeding electrons from the TCA cycle into the 'respiratory chain' – rather than NADH and succinate (Gnaiger 2020). This 'Complex II ambiguity' has deeply penetrated the scientific literature on bioenergetics without sufficient quality control. Therefore, a critical literature survey is needed to draw attention to widespread ambiguities, particularly in graphical representations of the mitochondrial electron transfer system, to ensure scientific standards in communications on bioenergetics.

2. Electron flow through CI and CII to the coenzyme Q junction

The reduced flavin groups FMNH_2 of flavin mononucleotide and FADH_2 of flavin adenine dinucleotide are at functionally comparable levels in the electron transfer through CI and CII, respectively (Figure 1a,b). FMNH_2 and FADH_2 are reoxidized downstream in CI and CII, respectively, by electron transfer converging at the Q-junction. The convergent architecture of the electron transfer system (ETS; in contrast to a linear electron transfer chain) is emphasized in Figures 1c and 1d (Hatefi 1962; Gnaiger 2020). Comparable to CII, several respiratory Complexes are localized in the mitochondrial inner membrane (mtIM) which catalyze electron transfer converging at the Q-junction, including electron transferring flavoprotein Complex (ETF) in fatty acid oxidation, glycerophosphate DH Complex (CGpDH), sulfide-ubiquinone oxidoreductase, choline DH, dihydro-orotate DH, and proline DH (Gnaiger 2020; Bénit et al 2022; Pallag et al 2022).

Complex II is a flavoprotein with a covalently bound flavin adenine dinucleotide as documented in early reports (Kearney 1960) and summarized in classical textbooks (Lehninger 1975; Tzagoloff 1982). Microscopic detail on the structure and function of CII (Cecchini 2003) has expanded our knowledge on the mechanism of enzyme assembly (Maklashina et al 2022), enzyme structure (Vercellino, Sazanov 2022; Karavaeva, Sousa

2023), kinetic regulation of CII activity (Mills et al 2018; Fink et al 2022), and associated pathologies (Bénit et al 2022; Iverson et al 2023).

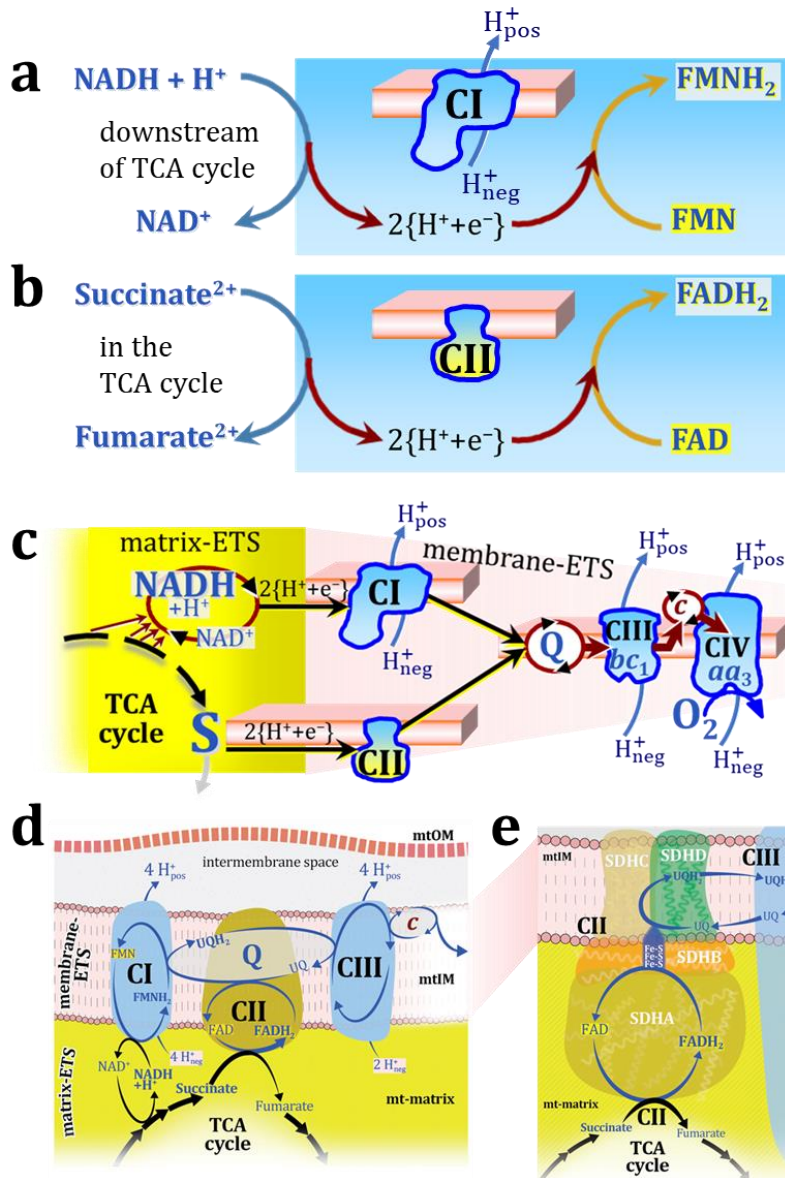


Figure 1. Complex II (SDH) bridges H⁺-linked electron transfer from the TCA cycle (matrix-ETS) to the electron transfer system (membrane-ETS) of the mt-inner membrane (mtIM). (a) NADH+H⁺ and (b) succinate are substrates of 2{H⁺+e⁻} transfer to the prosthetic groups FMN and FAD as the corresponding electron acceptors in CI and CII, respectively. (c) Symbolic representation of ETS pathway architecture. Electron flow converges at the N-junction (NAD⁺ → NADH+H⁺), and from NADH+H⁺ and succinate S at the Q-junction. CIII passes electrons to cytochrome c and in CIV to molecular O₂, 2{H⁺+e⁻}+0.5 O₂ → H₂O. (d) NADH+H⁺ and NAD⁺ cycle between matrix-dehydrogenases and CI, whereas FAD and FADH₂ cycle permanently bound within the same enzyme CII. Succinate and fumarate indicate the chemical entities

irrespective of ionization, whereas charges are shown in NADH (uncharged), NAD⁺, and H⁺. Joint pairs of half-circular arrows distinguish electron transfer 2{H⁺+e⁻} to CI and CII from vectorial H⁺ translocation across the mtIM (H⁺_{neg} → H⁺_{pos}). CI, CIII, and CIV pump hydrogen ions from the negatively (neg) to the positively charged compartment (pos). (e) Iconic representation of SDH subunits. SDHA catalyzes the oxidation succinate → fumarate + 2{H⁺+e⁻} and reduction FAD + 2{H⁺+e⁻} → FADH₂ in the soluble domain of CII. The iron-sulfur protein SDHB transfers electrons through Fe-S clusters to the mtIM domain where ubiquinone UQ is reduced to ubiquinol UQH₂ in SDHC and SDHD.

H⁺-linked two-electron transfer from succinate to flavin adenoside dinucleotide reduces the oxidized prosthetic group FAD to FADH₂ with formation of fumarate. This H⁺-linked electron transfer through CII is not coupled to H⁺ translocation across the mtIM. Hence, CII is not a H⁺ pump in contrast to the respiratory Complexes CI, CIII and CIV

through which electron transfer – more appropriately $2\{H^+e^-\}$ transfer (Table 1) – drives and maintains the protonmotive force.

The reversible oxidoreduction of succinate and fumarate is catalyzed in the soluble domain of CII extending from the mtIM into the mt-matrix. Succinate donates $2\{H^+e^-\}$ to FAD bound to the subunit SDHA which contains the catalytically active dicarboxylate binding site. The oxidized yellow (450 nm) form FAD functions as the hydrogen acceptor from succinate to the reduced internal product FADH₂ while fumarate is formed as the oxidized external product in the TCA cycle. In most flavin-linked dehydrogenases the flavin nucleotide is a tightly bound prosthetic group. In CII, however, it is even covalently and thus permanently bound to the enzyme during the catalytic cycle when the redox state is regenerated in each enzymatic turnover. FADH₂ relays electrons further through a series of iron-sulfur redox centers in SDHB to ubiquinone in the membrane domain harboring SDHC and SDHD (Moosavi et al 2019) (Figure 1e).

Simple arrows (Figure 1a-c) or pairs of rounded arrows – an external arrow touching the enzyme and an internal arrow within the enzyme – indicate H⁺-linked electron transfer (Hsu et al 2022) (Figure 1d,e). Caution is warranted to distinguish three types of transformation with hydrogen ions: (1) acid-base reactions equilibrating fast without catalyst, (2) slow catalyst-dependent redox transfer of hydrogen atoms $2\{H^+e^-\}$, and (3) vectorial hydrogen ion translocation through H⁺ pumps (Table 1).

Table 1. Three distinct types of transformation with hydrogen ions H⁺.

Transformation	Equation	Type
1. acid-base equilibrium	$H_3O^+ \leftrightarrow H_2O + H^+$ $H_2CO_3 \leftrightarrow HCO_3^- + H^+$	(a) scalar, (b) chemical, fast
2a. H ⁺ -linked electron transfer, oxidation	$Malate^{2-} \rightarrow Oxaloacetate^{2-} + 2\{H^+e^-\}$ $Succinate^{2-} \rightarrow Fumarate^{2-} + 2\{H^+e^-\}$	(c) scalar, (d) chemical, slow
2b. H ⁺ -linked electron transfer, reduction	$2\{H^+e^-\} + NAD^+ \rightarrow NADH + H^+$ $2\{H^+e^-\} + E-FAD \rightarrow E-FADH_2$	(e) scalar, (f) chemical, slow
3. transport, translocation	pumping: $H^+_{neg} \rightarrow H^+_{pos}$ diffusion: $H^+_{pos} \rightarrow H^+_{neg}$	(g) vectorial, (h) compartmental, transmembrane

3. The source and consequence of Complex II ambiguities

'No representation is ever perfectly expressive, for if it were it would not be a representation but the thing itself' (Grosholz 2007).

Ambiguities emerge if the representation of a concept is vague to an extent that allows for equivocal interpretations. As a consequence, even a basically clear concept (Figure 1) may be communicated as a divergence from an established 'truth'. The comparison between NADH linked to CI and FADH₂ (instead of succinate) linked to CII leads us astray, as illustrated by the following textbook-quotes (Cooper 2000) (Figure 2).

(1) *'Electrons from NADH enter the electron transport chain in complex I, .. A distinct protein complex (complex II), which consists of four polypeptides, receives electrons from the*

citric acid cycle intermediate, succinate (Figure 10.9). These electrons are transferred to $FADH_2$, rather than to $NADH$, and then to coenzyme Q.' Note the suggestive comparison of $FADH_2$ and $NADH$.

(2) 'In contrast to the transfer of electrons from $NADH$ to coenzyme Q at complex I, the transfer of electrons from $FADH_2$ to coenzyme Q is not associated with a significant decrease in free energy and, therefore, is not coupled to ATP synthesis.' Note that CI catalyzes electron transfer from $NADH$ to coenzyme Q. In contrast, electron transfer from $FADH_2$ to coenzyme Q is downstream of succinate oxidation by CII. Thus instead of the Gibbs force ('decrease in free energy') in $FADH_2 \rightarrow Q$, the total Gibbs force (Gnaiger 2020) in $S \rightarrow FADH_2 \rightarrow Q$ must be accounted for. (In parentheses: Redox-driven proton translocation must be distinguished from phosphorylation of ADP driven by the protonmotive force).

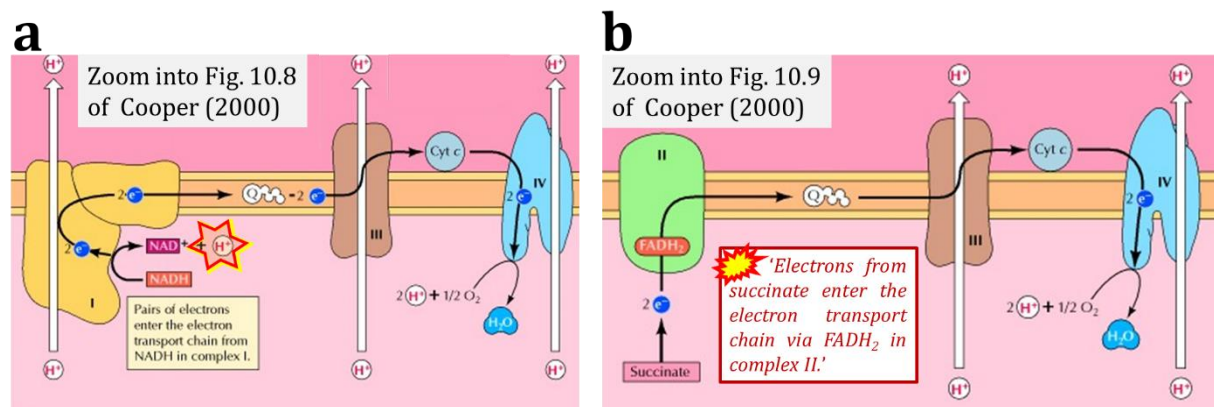


Figure 2. Electron transfer to CI and CII. Zoom into figures of Cooper (2000). **(a)** The marked H^+ is consumed in H^+ -linked electron transfer instead of being produced. **(b)** Marked quote inserted from the legend to Fig. 10.9.

(3) '.. electrons from succinate enter the electron transport chain via $FADH_2$ in complex II.' Note that CII receives electrons from succinate via FAD. The ambiguity is caused by a lack of unequivocal definition of the electron transfer system ('electron transport chain'; Supplement 1). Two contrasting definitions are implied of the 'electron transport chain' or ETS. (a) CII is part of the ETS. Hence electrons enter the ETS in the succinate branch from succinate but not from $FADH_2$ – from the matrix-ETS to the membrane-ETS (Figure 1c,d). (b) If electrons enter the 'electron transport chain via $FADH_2$ in complex II', then subunit SDHA would be upstream and hence not part of the ETS (to which conclusion obviously nobody would agree). There remains the ambiguity of electron entry into CII from succinate (Figure 1) or from $FADH_2$ as the 'product' of succinate dehydrogenase in the TCA cycle (Figure 3a,b).

4. The $FADH_2$ - FAD confusion in the succinate-pathway

'Like drops of water on stone, one drop will do no harm, but over time, grooves are cut deep' (Wardle 2023).

The narrative that the reduced cofactors $NADH$ and $FADH_2$ feed electrons from the TCA cycle into the mitochondrial electron transfer system causes confusion. As a consequence, the prosthetic group $FADH_2$ appears erroneously as the substrate of CII in the ETS linked to succinate oxidation. This error is widely propagated in 99 publications found from 2001 to 2023 (Supplements 2 to 6) and numerous educational websites

(Supplement 7). Clarification is required (Gnaiger 2020; page 48). The following examples illustrate the transition from ambiguity to erroneous representation.

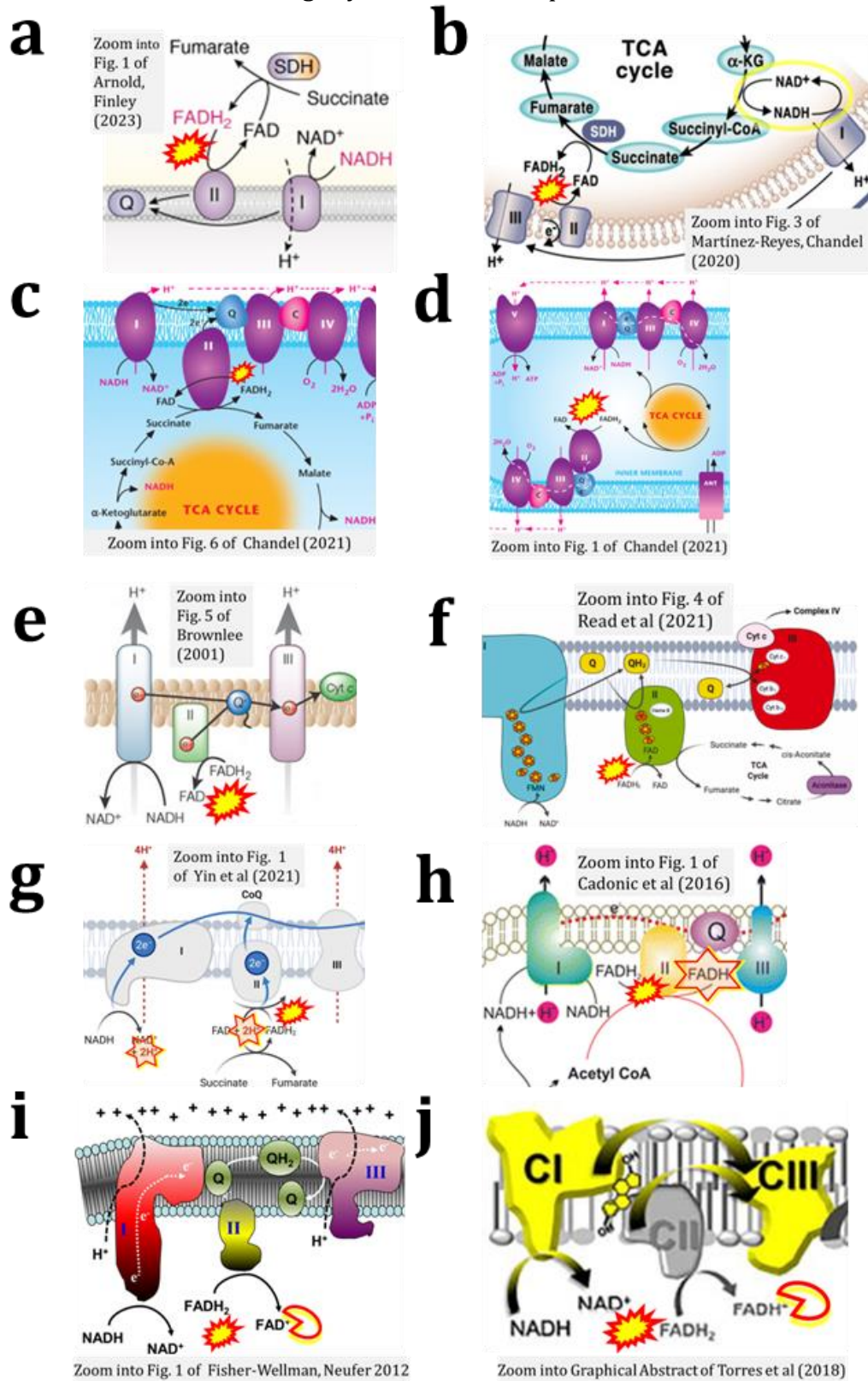


Figure 3. Complex II ambiguities. FADH₂ depicted as product and substrate of Complex II by (a) Arnold, Finley (2023), (b) Martínez-Reyes, Chandel (2020). NADH and NAD⁺ cycle between different types of enzymes (yellow circle), in contrast to the FADH₂-FAD

cycle located within the same enzyme (SDH and CII are synonyms). **(c)** From ambiguity to **(d)** graphical misconception (Chandel 2021). **(e)** FADH₂ shown as substrate of CII (Brownlee 2001). **(f)** FAD shown as product in the mt-matrix and inside CII (Read et al 2021). **(g)** Unjustified indication of 2H⁺ formation in the mt-matrix (Yin et al 2021), not trivial considering the concept of the protonmotive force. **(h)** FADH₂ as substrate of CII and FADH (?) as product (Cadonic et al 2016). **(i)** The NADH→NAD⁺ analogy is taken to the level of copying a charge to FAD⁺ (Fisher-Wellman, Neuffer 2012) or **(j)** FADH⁺ (Torres et al 2018).

(1) Ambiguities appear in graphical representations, where FADH₂ is the product of SDH and the substrate of CII – synonymous with SDH (Figure 3a,b; Suppl Figure S2).

(2) Ambiguity evolved to misconception in graphical representations (Figure 3c,d).

(3) Graphical errors on electron entry from FADH₂ into CII show up without comment (Figure 3e,f; Suppl Figures S3).

(4) Instead of NADH+H⁺→NAD⁺ there appears NADH→NAD⁺+H⁺ (or +2H⁺) and by analogy FADH₂→FAD +2H⁺ (Figure 3g; Suppl Figure S4) or FADH₂→FADH (Figure 3h). The analogy NADH→NAD⁺ is taken further to include a charge for FAD or even writing FADH⁺ (Figure 3i,j; Suppl Figures S5 and S6). Disturbing patterns are shown in various figures with analogous representations of oxidation of NADH and FADH₂ (Table 2).

Finally, error propagation from graphical representation (Figure 3) leads to misconception in the text: 'SDH reduces FAD to FADH₂, which donates its electrons to complex II'; 'each complete turn of the TCA cycle generates three NADH and one FADH₂ molecules, which donate their electrons to complex I and complex II, respectively'; 'complex I and complex II oxidize NADH and FADH₂, respectively' (Arnold, Finley 2023).

Table 2. Misconceptions in graphical representations of electron entry into CII.

Analogy with NADH	Suppl Figure	FADH ₂	Suppl Figure
NADH + H ⁺ → NAD ⁺			
NADH → NAD ⁺ + H ⁺	S3d,q, κ, o, v	FADH ₂ → FAD	S2, S3
NADH → NAD ⁺ + H ⁺	S4a,e,g	FADH ₂ → FAD + 2H ⁺	S4a-i
NADH → NAD ⁺ + 2H ⁺	S4c,f,h,i		
		FADH ₂ → FAD ⁺	S5a-i
NADH + H ⁺ → NADH	S6a	FADH ₂ → FADH	S6a-d
		FADH ₂ → FADH ⁺	S6e
		FADH →	S6f
NADH → NAD + H ⁺	S4b	FADH → FAD ⁺	S6g
NADH → NAD ⁺ + H ⁺	S6h	FADH → FAD ⁺ + H ⁺	S6h
NADH → NAD ⁺ + H ⁺	S6i	FADH → FAD ⁺ + 2H ⁺	S6i

Electron transfer from succinate in the TCA cycle to the prosthetic group FAD is a redox reaction, where oxidation (ox) of succinate yields 2{H⁺+e⁻} – two hydrogen ions and two electrons – which are donated in the reduction (red) of FAD to FADH₂ (Table 1),



The net redox reaction equation is



Commonly the charges of succinate, fumarate (Eq. 1), and other metabolites are not shown explicitly to simplify graphical representations of metabolic pathways. But NAD^+ is clearly distinguished from FAD (Figure 1). In $2\{\text{H}^++\text{e}^-\} + \text{NAD}^+ \rightarrow \text{NADH}+\text{H}^+$ the final H^+ is frequently omitted (Figure 3). One hydrogen atom is transferred directly from the hydrogen donor (e.g. malate) to NAD^+ without dilution by the aqueous H^+ whereas the other forms an aqueous hydrogen ion (Lehninger 1975). The equilibrium (Eq. *e* in Table 1) depends on pH. In contrast, Eq. 1b (Eq. *f* in Table 1) is independent of pH. The fundamental difference between H^+ and $2\{\text{H}^++\text{e}^-\}$ in Eq. *e* (Table 1) is lost in representations such as Figure 3.

In summary, two-electron oxidation of succinate is redox-linked to reduction of FAD to FADH_2 . In terms of electron entry into CII many publications show it in the wrong direction, i.e. FADH_2 as electron donor from the TCA cycle to CII (Figure 3; Suppl Figures S2 to S6). This erroneous presentation has a logical consequence. β -oxidation generates FADH_2 (Figure 4). If FADH_2 would donate electrons to CII, then CII can be seen as an enzyme involved downstream of FADH_2 in FAO. This is incorrect as clarified in the next section.

5. Complex II and fatty acid oxidation

Electron transferring flavoprotein CETF and CI are the respiratory Complexes involved in convergent electron entry into the Q-junction during FAO (Figure 4).

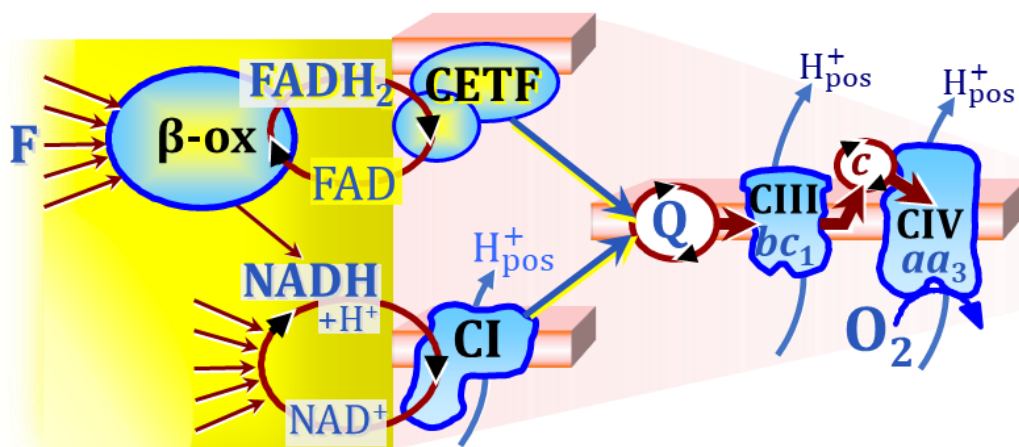


Figure 4. Fatty acid oxidation through the β -oxidation cycle (β -ox), the multi-enzyme electron transferring flavoprotein Complex (CETF, ETF:ETFDH; see text), and Complex I (CI) with convergent electron transfer into the Q-junction.

In the β -oxidation cycle of FAO, acetyl-CoA and the reducing equivalents FADH_2 and NADH are formed in reactions catalyzed by acyl-CoA dehydrogenases and hydroxyacyl-CoA dehydrogenases, respectively, in the mitochondrial matrix (Houten et al 2016). When FADH_2 is erroneously shown free floating in the mt-matrix as a substrate of CII, a dubious role of CII in FAO is suggested as a consequence (Figure 5; Supplement 8).

Lemmi et al (1990) noted: ‘mitochondrial Complex II also participates in the oxidation of fatty acids’. This holds for the oxidation of acetyl-CoA in the TCA cycle, forming NADH and succinate with downstream electron flow through CI and CII, respectively, into

the Q-junction (Figure 1). In contrast, electron transfer from FADH₂ formed during β -oxidation proceeds through electron transferring flavoprotein ETF. Fatty acyl-CoA dehydrogenases in the mitochondrial matrix reduce FAD to FADH₂. The FADH₂ of the fatty acyl-CoA DHs is reoxidized by the FAD-containing ETF (Crane, Beinert 1956).

ETF and ETFDH (Wang et al 2019; or electron transfer flavoprotein:ubiquinone oxidoreductase ETF-QO, Watmough, Frerman 2010) comprise the ETF Complex (CETF), i.e. the ETF:ETFDH or ETF:ETF-QO system. CETF links electron transfer in β -oxidation to electron entry into the Q-junction independent of CII (Figure 4). Thus FADH₂ can be seen as an internal substrate of CETF, comparable to the external substrates NADH for CI, succinate for CII, and glycerophosphate for CGpDH.

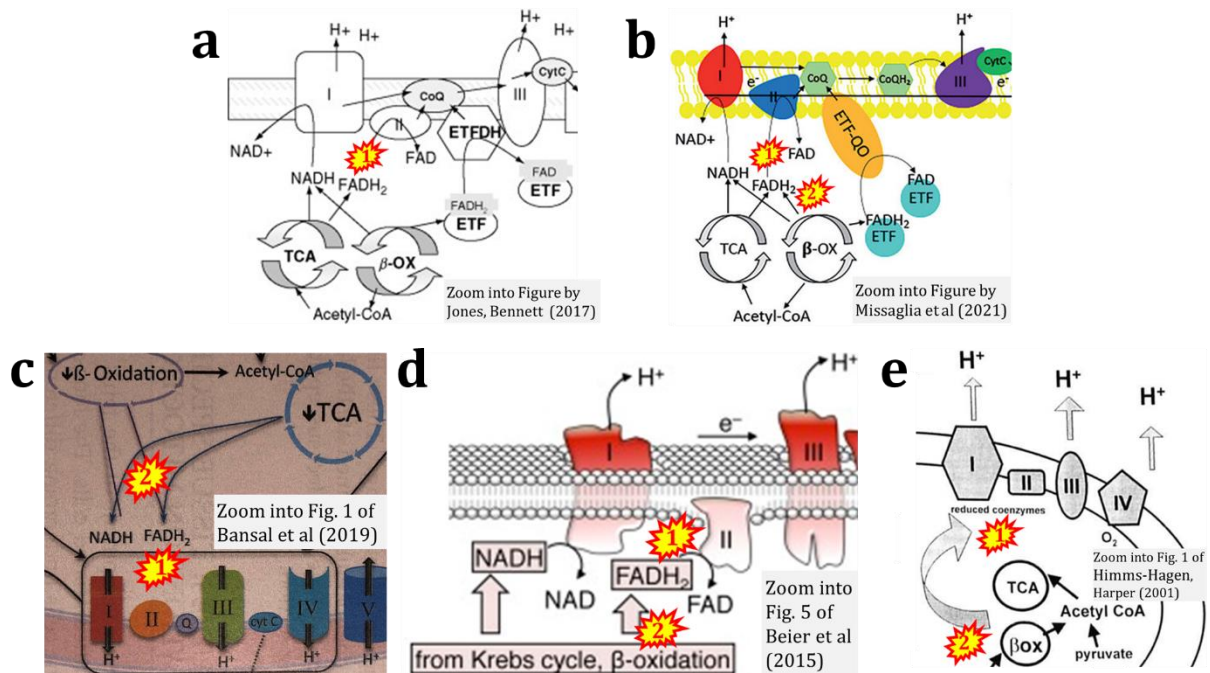


Figure 5. When FADH₂ is erroneously shown as a substrate of CII (1), a role of CII in oxidation of FADH₂ from fatty acid oxidation is suggested as a consequence (2). Zoom into figures from (a) Jones, Bennett (2017); (b) Missaglia et al (2021); (c) Bansal et al (2019); (d) Beier et al (2015); (e) Himms-Hagen, Harper (2001).

6. Conclusions

There is currently ambiguity surrounding the precise role of Complex II in fatty acid oxidation. While Complex II is not essential for fatty acid oxidation, it plays a regulatory role by sensing changes in metabolic demand and activating the TCA cycle for oxidation of acetyl-CoA depending on the metabolic conditions. This regulatory function may be particularly important during periods of low oxygen availability or high energy demand. The integration of FAO with the membrane-bound ETS (Wang et al 2019) has significant implications for understanding and treating disorders related to β -oxidation and oxidative phosphorylation. Using precisely defined terminology can prevent misunderstandings (Gnaiger et al 2020; footnotes in Supplement 1). Do misinformed diagrams – from ambiguous electron transfer (Suppl Figures S2 to S8) to presentation of CII as a H⁺ pump (Suppl Figure S9) – cast any doubts on the quality of the publication? Whether using iconic or symbolic elements in graphical representations, incorporating

complementary text not only enhances the communication of intended meaning but diagrams will be improved in the process.

When peer review provides insufficient help for corrections, post-peer review by editors and critical readers is required for revisions of articles which may be re-published as living communications (Gnaiger 2021). The present review aims to raise awareness in the scientific community about the inherent ambiguity crisis, complementary to addressing the widely recognized issues of the reproducibility and credibility crisis (Gall et al 2017). The term 'crisis' is rooted etymologically in the Greek word *krinein*: meaning to 'separate, decide, judge'. In this sense, science and communication in general are a continuous crisis at the edge of separating clarity or certainty from confusing double meaning to fake-news. Reproducibility relates to the condition of repeating and confirming calculations or experiments presented in a published resource. While ambiguity is linked to relevant issues of reproducibility, it extends to the communications space of terminological and graphical representations of concepts (Grosholz 2007). Type 1 ambiguities are the inevitable consequence of conceptual evolution, in the process of which ambiguities are replaced by experimentally and theoretically supported paradigm shifts to clear-cut theorems. In contrast, type 2 ambiguities are traced in publications that reflect merely a disregard and ignorance of established concepts without an attempt to justify the inherent deviations from high-quality science. There are many shades of grey between these types of ambiguity.

A prominent case of ambiguity in the grey zone between types 1 and 2 has been uniquely demonstrated by analysis of the popular notion of 'oxidative stress' - a term more frequently found in PubMed than 'mitochondria', widely used with vage definition and without expression by numerical values and corresponding units (Azzi 2022). Another example closer to type 2 ambiguity is the use of the terms and experimental application of 'hypoxia' and 'normoxia' in bioenergetics, when air-level normoxic conditions for isolated mitochondria and cultured cells are effectively hyperoxic and may cause oxidative damage (Gnaiger et al 2000; Donnelly et al 2022). Another ambiguity in bioenergetics links to the confusing use of the terms uncoupling, decoupling, dyscoupling, where rigorous definition is warranted (Gnaiger et al 2020). Linking bioenergetics to physical chemistry and the thermodynamics of irreversible processes, the ambiguous use (type 1) of the terms force and pressure (vant'Hoff 1901; Einstein 1905; Nernst 1921; Prigogine 1967; Mitchell 1966) has deep consequences on the enigmatic concept of non-ohmic flux-force relationships in the context of mitochondrial membrane potential and the protonmotive force (Gnaiger 2020).

The present review adds Complex II ambiguities to the growing list. Clarification instead of perpetuation of Complex II ambiguities leads to a better representation of fundamental concepts of bioenergetics and helps to maintain the high scientific standards required for translating knowledge on metabolism into clinical solutions for mitochondrial diseases.

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Abbreviations

2{H ⁺ +e ⁻ }	redox equivalents in H ⁺ -linked electron transfer	mt-matrix	mitochondrial matrix
CI	Complex I	mtIM	mitochondrial inner membrane
CII	Complex II	NADH ₂	reduced nicotinamide adenine dinucleotide
CETF	electron transferring flavoprotein Complex (ETF:ETFDH)	NAD ⁺	oxidized nicotinamide adenine dinucleotide
CGpDH	mt-glycerophosphate dehydrogenase Complex	Q	ETS-reactive coenzyme Q, oxidation state is not implied
DH	dehydrogenase	QFR	mena-quinol-fumarate oxidoreductase
FADH ₂	reduced flavin adenoside dinucleotide	SQR	succinate-ubiquinone oxidoreductase
FAD	oxidized flavin adenoside dinucleotide	SDH, SDHABCD	succinate dehydrogenase, CII tricarboxylic acid cycle
FAO	fatty acid oxidation	TCA cycle	
FMNH ₂	reduced flavin mononucleotide		

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Supplement 1. Footnotes on terminology

Coenzyme: ‘The dissociable, low-relative-molecular-mass active group of an enzyme which transfers chemical groups, hydrogen, or electrons. A coenzyme binds with its associated protein (apoenzyme) to form the active enzyme (holoenzyme) (Burtis, Geary 1994). ‘A low-molecular-weight, non-protein organic compound participating in enzymatic reactions as dissociable acceptor or donor of chemical groups or electrons’ - <https://www.ebi.ac.uk/chebi/searchId.do?chebiId=CHEBI:23354> (CHEBI:23354, retrieved 2023-06-21). A coenzyme or cosubstrate is a cofactor that is attached loosely and transiently to an enzyme. NADH is listed as a coenzyme, which should be regarded as a substrate of pyridine-linked dehydrogenases (Lehninger 1975).

Cofactor: A cofactor is ‘an organic molecule or ion (usually a metal ion) that is required by an enzyme for its activity. It may be attached either loosely (coenzyme) or tightly (prosthetic group)’ - <https://www.ebi.ac.uk/chebi/searchId.do?chebiId=23357> (CHEBI:23357, retrieved 2023-06-21).

Electron transfer system ETS: The *convergent* architecture of the electron transfer *system* is emphasized in contrast to *linear* electron transfer *chains* ETCs within segments of the ETS.

Electron transfer: A distinction is necessary between electron *transfer* in redox reactions and electron *transport* (translocation) in the diffusion of charged ionic species within or between cellular compartments. The symbol $2\{H^+e^-\}$ is introduced to indicate H⁺-linked electron transfer of two hydrogen ions and two electrons in a redox reaction.

Gibbs force: In contrast to the extensive quantity Gibbs energy [J], Gibbs force [J·mol⁻¹] is an intensive quantity expressed as the partial derivative of Gibbs energy [J] per advancement of a reaction [mol] (Gnaiger 1993; 2020).

H⁺-linked electron transfer: The term H⁺-coupled electron transfer (Hsu et al 2022) is replaced by H⁺-*linked* electron transfer, to avoid confusion with *coupled* H⁺ translocation.

Matrix-ETS: Electron transfer and corresponding OXPHOS capacities are classically studied in mitochondrial preparations as oxygen consumption supported by various fuel substrates undergoing partial oxidation in the mt-matrix, such as pyruvate, malate, succinate, and others. Therefore, the *matrix* component of ETS (matrix-ETS) is distinguished from the ETS *bound to the mt-inner membrane* (membrane-ETS; Gnaiger et al 2020).

Membrane-ETS: Electron transfer is frequently considered as the segment of redox reactions linked to the mtIM. However, the *membrane*-ETS is only part of the total ETS, which includes the upstream *matrix*-ETS.

Misinformation: Misinformation is the mistaken sharing of the same content (Wardle 2023).

Prosthetic group: A prosthetic group is a cofactor that is ‘a tightly bound, specific nonpolypeptide unit in a protein determining and involved in its biological activity’ - <https://www.ebi.ac.uk/chebi/searchId.do?chebiId=CHEBI:26348> (CHEBI:26348, retrieved 2023-06-21). A prosthetic group is attached permanently and tightly or

even covalently to an enzyme and that is regenerated in each enzymatic turnover. FAD is the prosthetic group of flavin-linked dehydrogenases, covalently bound to CII.

Substrate: A substrate in a chemical reaction has a negative stoichiometric number since it is consumed, whereas a product has a positive stoichiometric number since it is produced. The general definition of a substrate in an enzyme-catalyzed reaction relies on the definition of the chemical reaction, without restriction to the nature of the substrate, i.e. independent of the substrate being a chemical entity in solution or a loosely bound cosubstrate (coenzyme) or even a tightly bound prosthetic group. The latter may be explicitly distinguished as a bound (internal) substrate from a free (external) substrate. Even different substrate pools may coexist (e.g. CoQ).

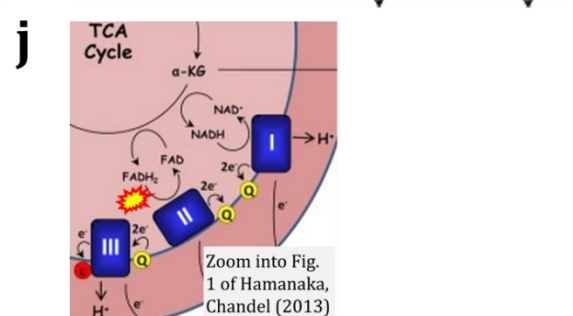
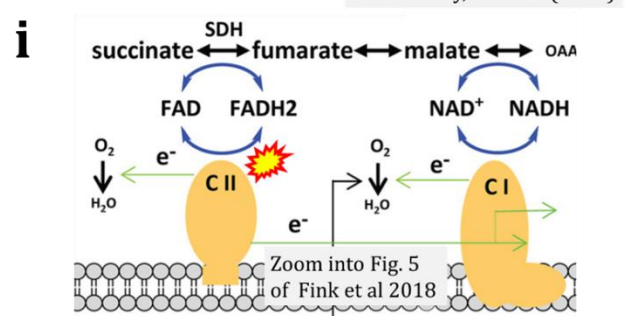
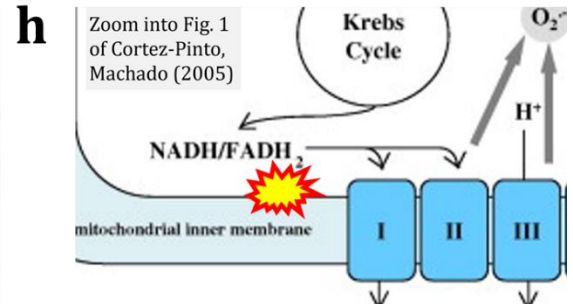
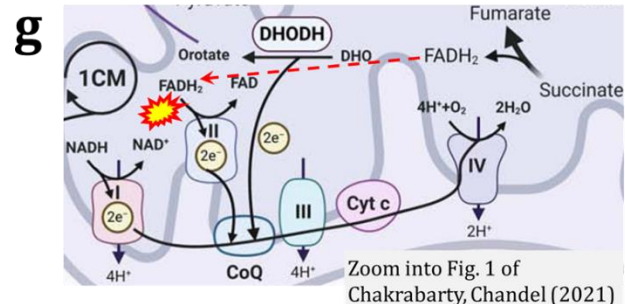
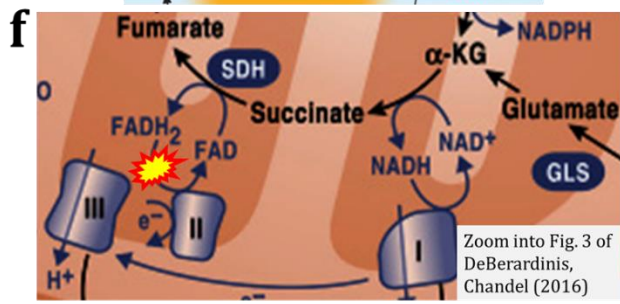
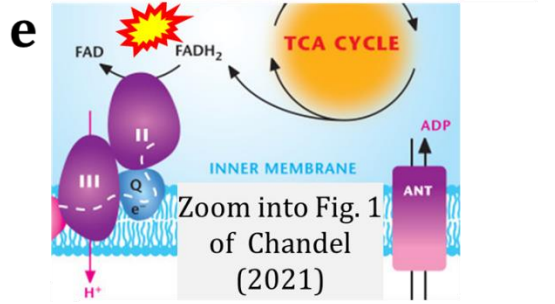
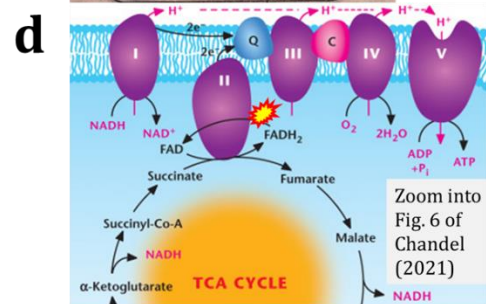
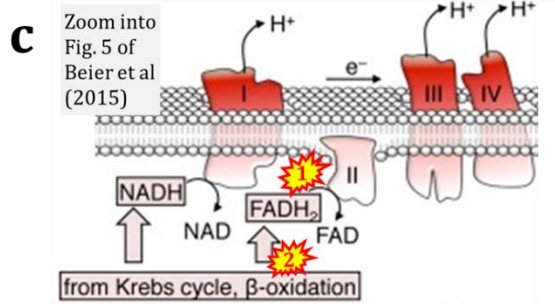
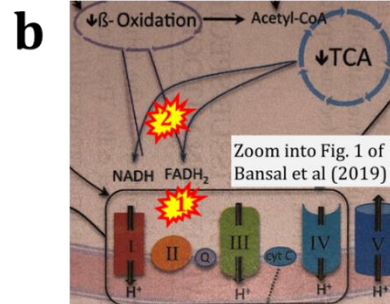
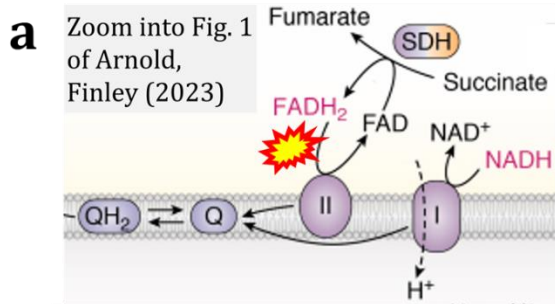
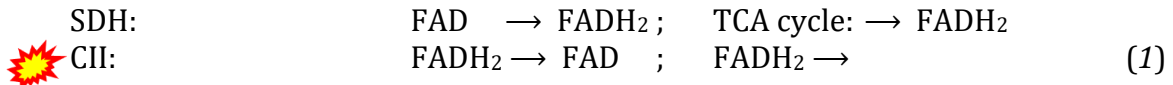
2{H⁺+e⁻}: In H⁺-linked two-electron transfer, 2H⁺ + 2e⁻, 'the terms reducing equivalents or electron equivalents are used to refer to electrons and/or hydrogen atoms participating in oxidoreductions' (Lehninger 1975). The symbol 2[H] is frequently used to distinguish reducing equivalents in the transfer from hydrogen donors to hydrogen acceptors from aqueous H⁺. Acid-base reactions obtain equilibrium fast without catalyst, whereas the slow oxidation-reduction reactions require an enzyme to proceed. However, 2[H] does not explicitly express that it applies to both *electron* and *hydrogen ion* transfer. Brackets are avoided to exclude the confusion with amount-of-substance concentrations frequently indicated by brackets. H⁺-linked two-electron transfer 2{H⁺+e⁻} is distinguished from single-electron transfer {H⁺}+{e⁻}.

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Supplement 2

FAD a substrate of SDH and FADH₂ a substrate of CII (Figure S2)



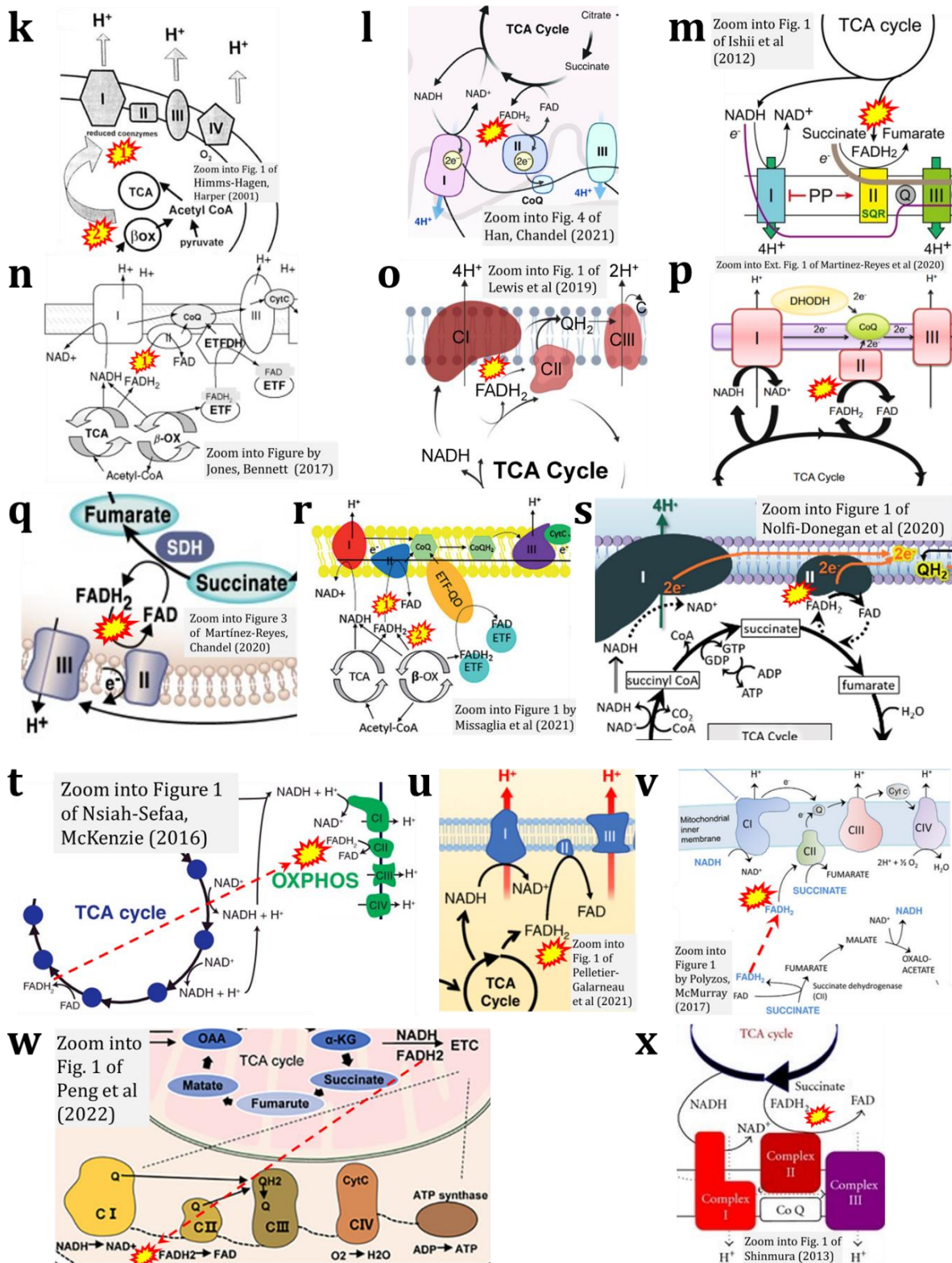


Figure S2. Complex II ambiguities in graphical representations on FADH₂ as a substrate of Complex II in the canonical forward electron transfer. The TCA cycle reduces FAD to FADH₂ – in several cases shown to be catalyzed by SDH. Then FADH₂ is erroneously shown to feed electrons into CII. Alphabetical sequence of publications from 2001 to 2023. See References for Figure S2.

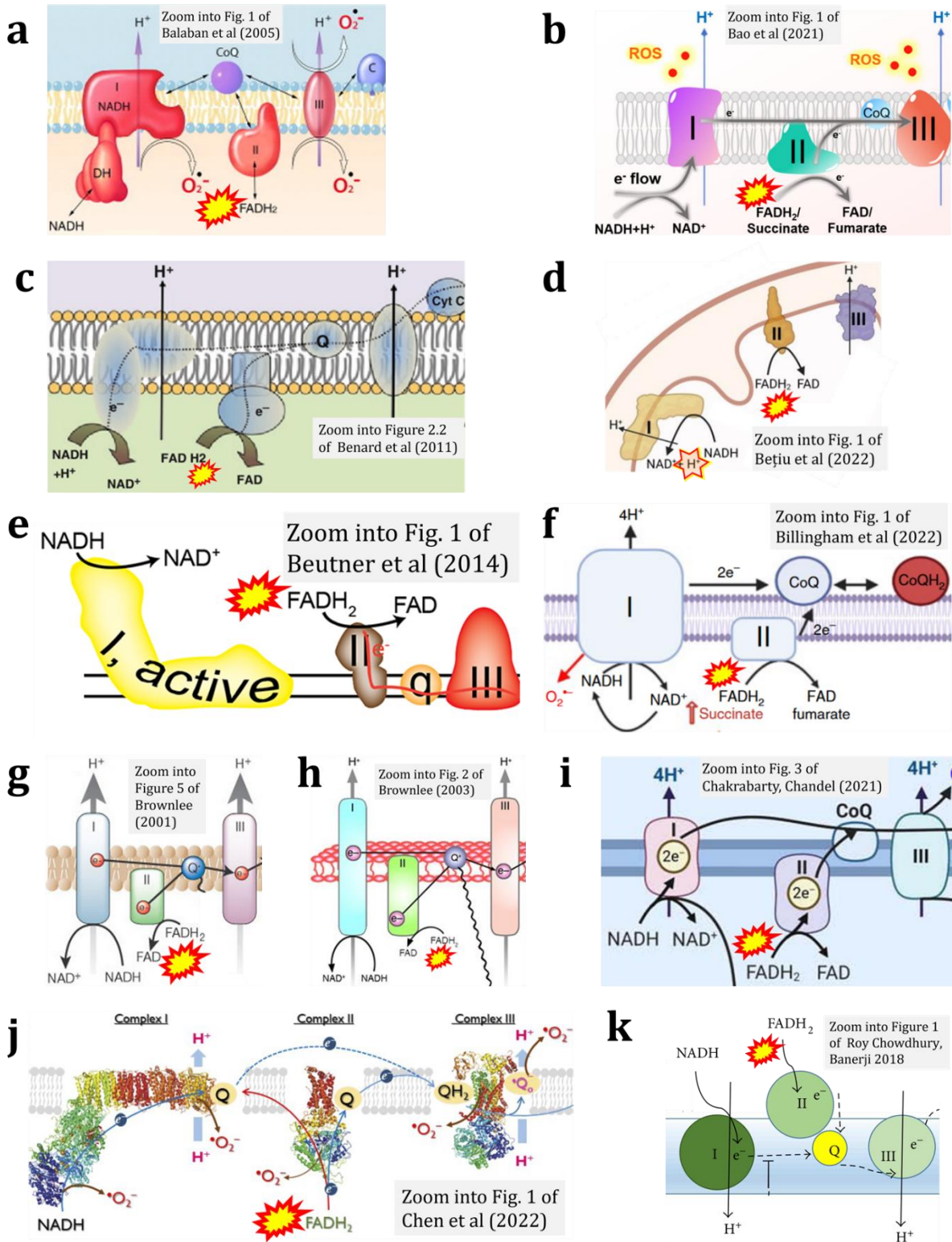
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Supplement 3

FADH₂ as substrate of CII (Figure S3)



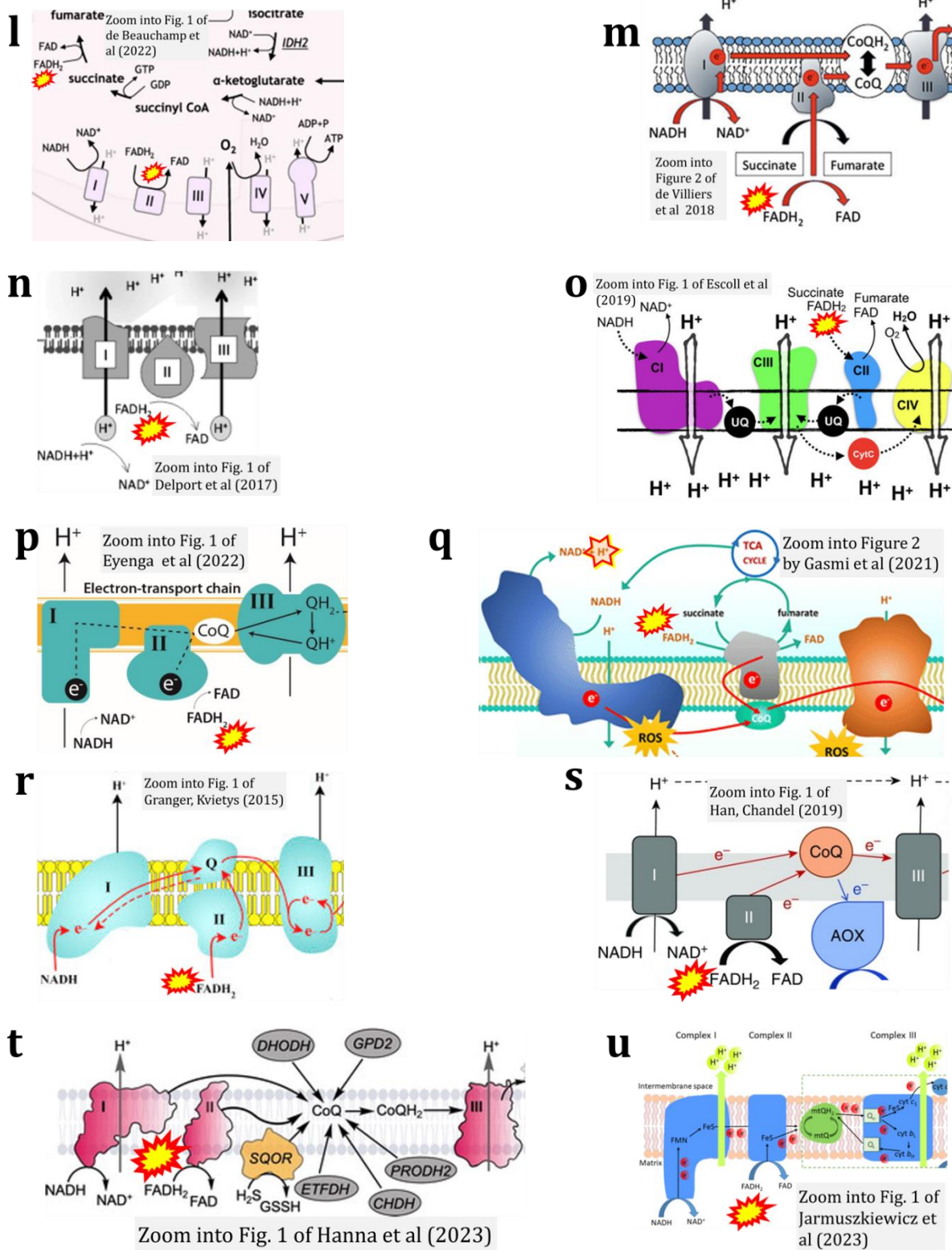


Figure S3. Continued

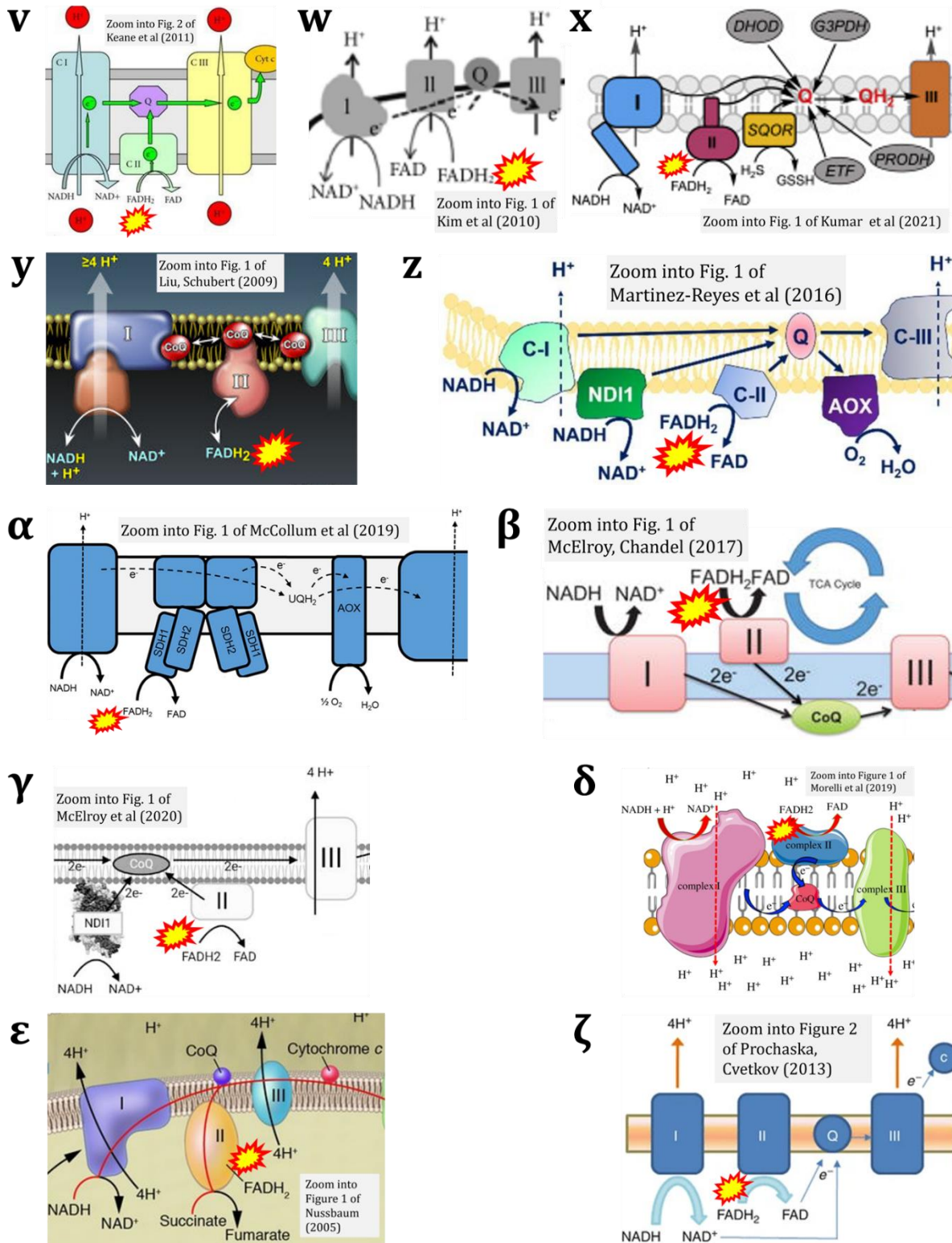


Figure S3. Continued

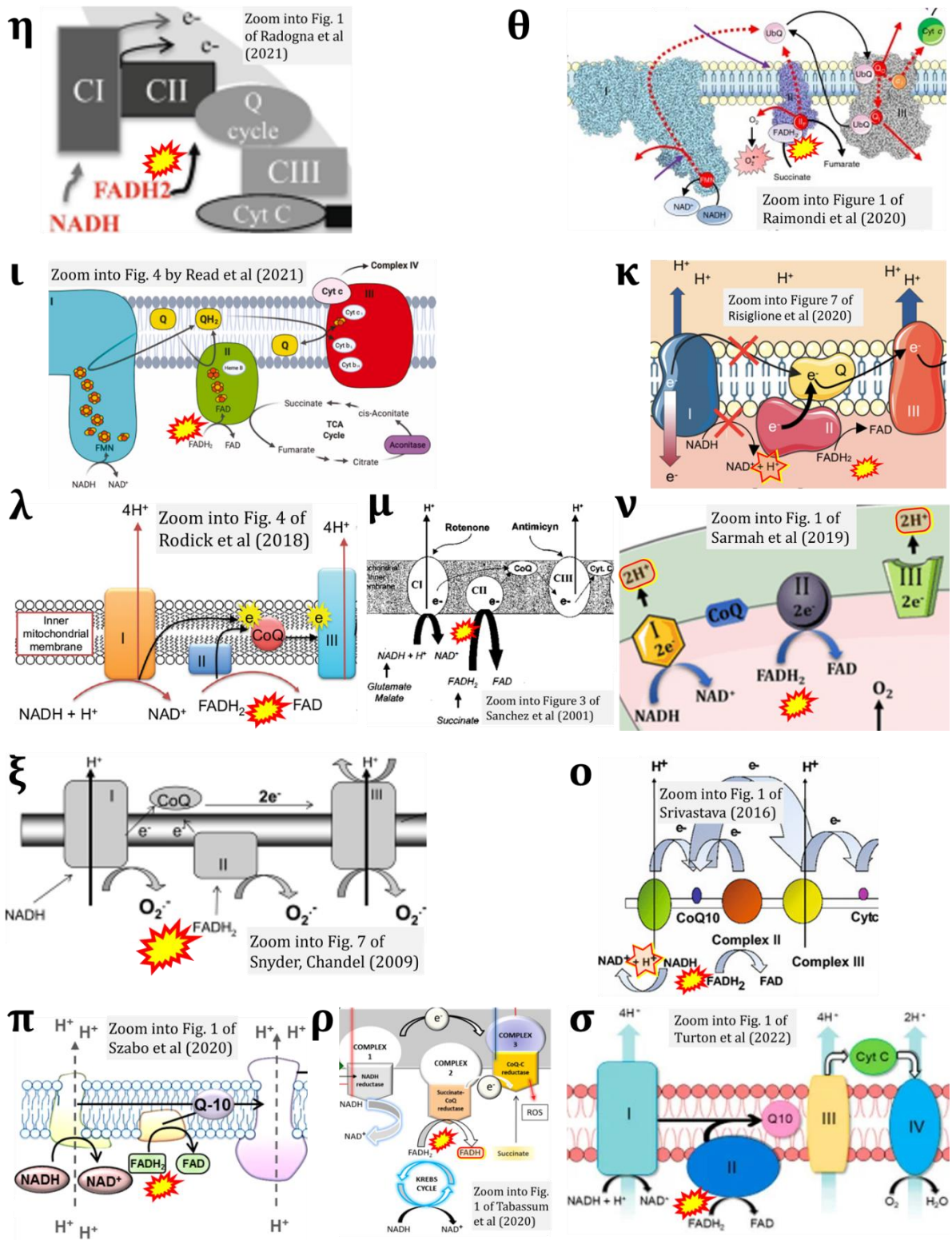


Figure S3. Continued

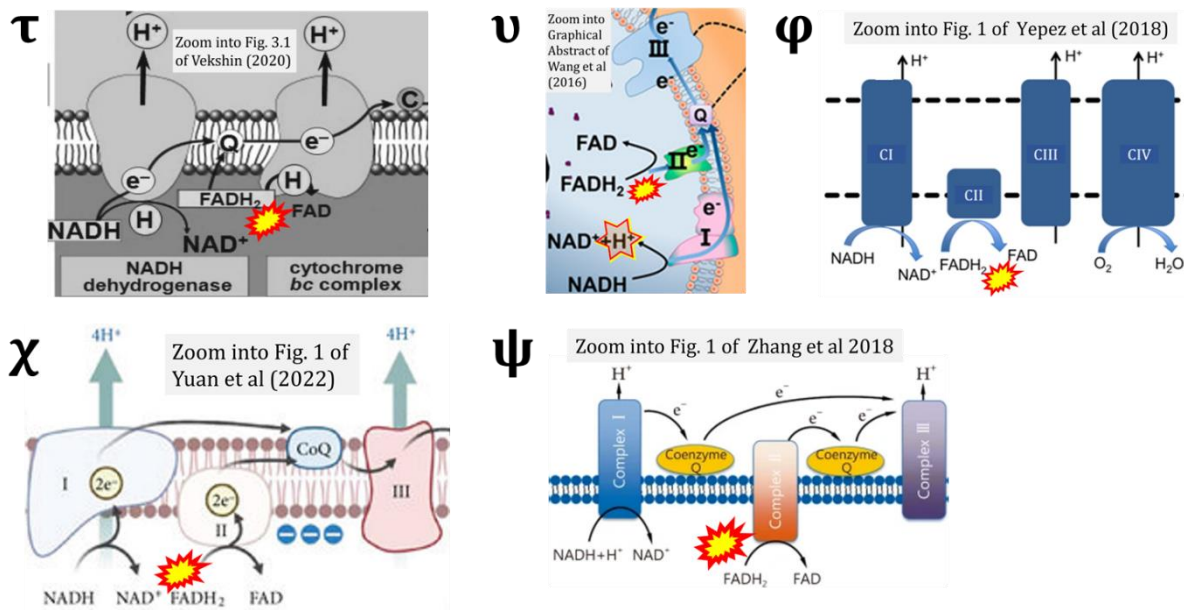


Figure S3. Complex II ambiguities in graphical representations on FADH₂ as a substrate of Complex II in the canonical forward electron transfer. Alphabetical sequence of publications from 2001 to 2023. See References for Figure S3.

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Supplement 4

FADH₂ as substrate of CII and FAD + 2H⁺ as products (Figure S4)

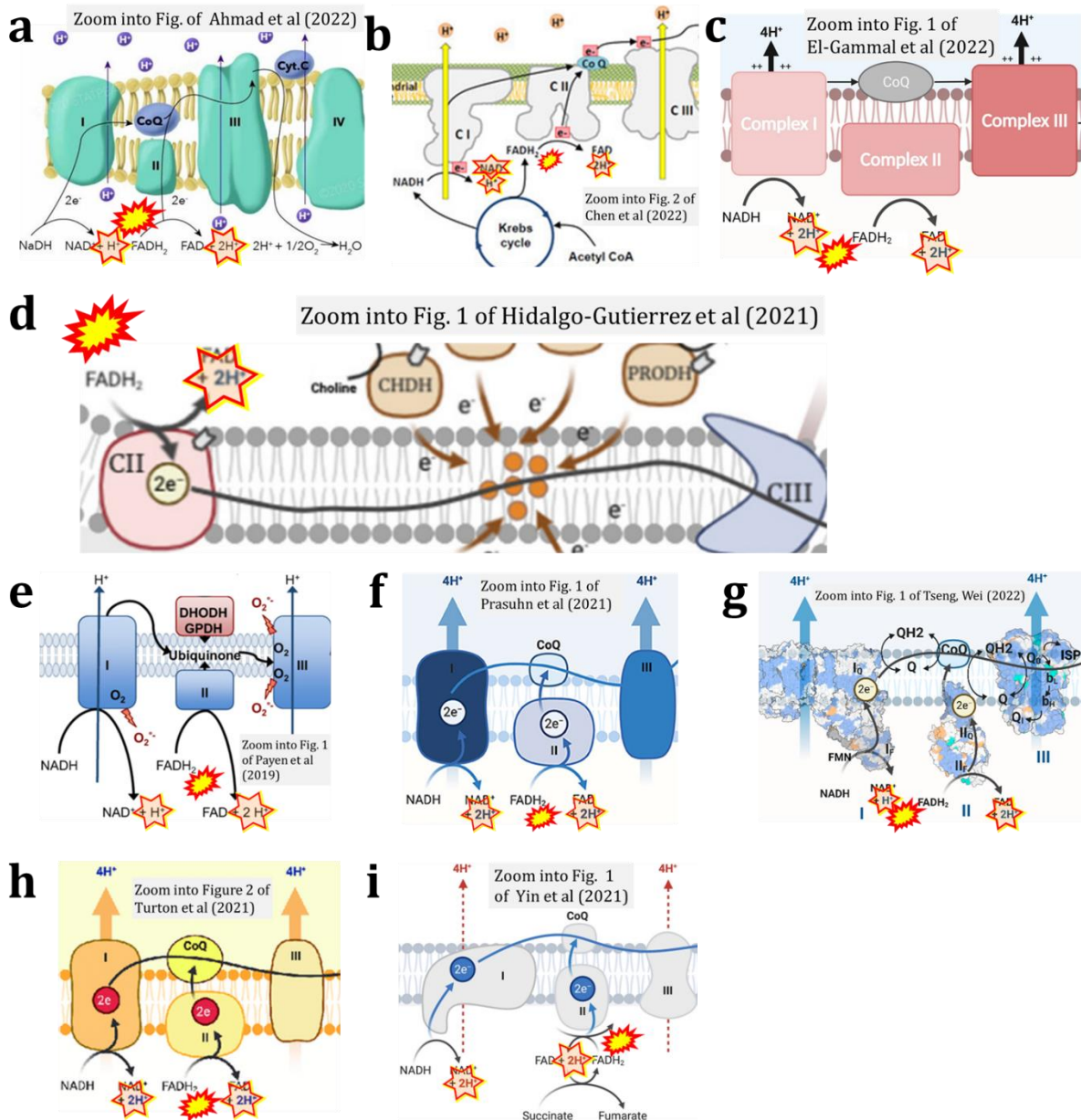
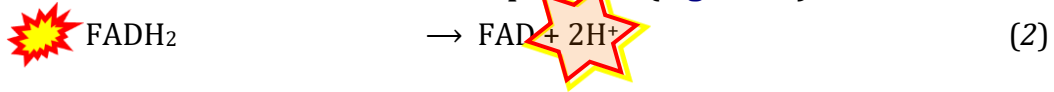


Figure S4. Complex II ambiguities: FADH₂ as substrate of CII and FAD + 2H⁺ as products. Alphabetical sequence of publications from 2001 to 2023. See References for Figure S4.

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Supplement 5

FADH₂ as substrate of CII and FAD⁺ as product (Figure S5)

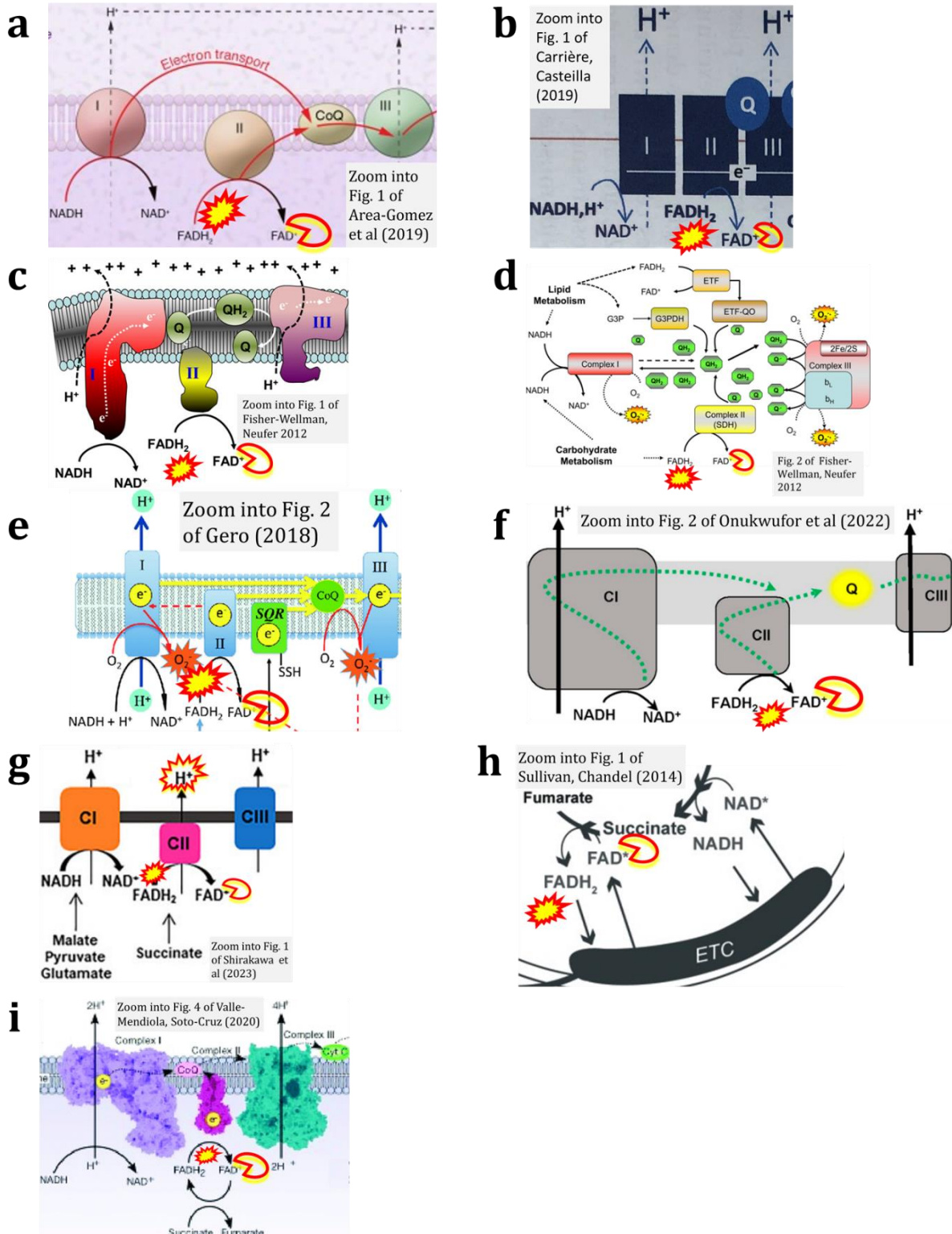










Figure S5. Complex II ambiguities: FADH₂ as substrate of CII and FAD⁺ as product. Alphabetical sequence of publications from 2001 to 2023. See References for Figure S5.

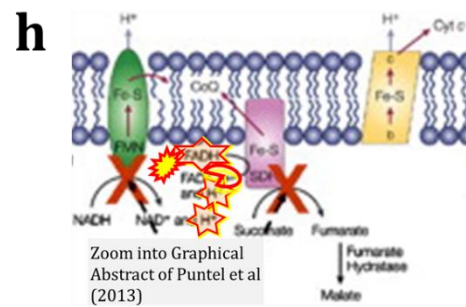
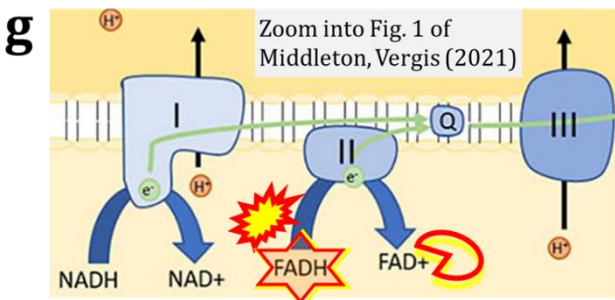
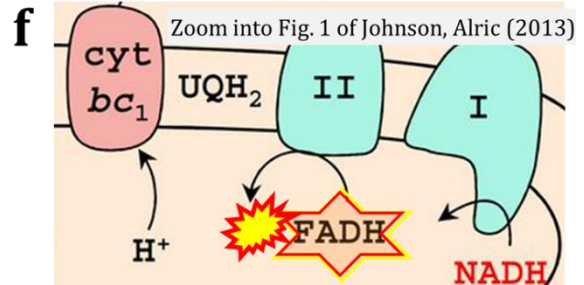
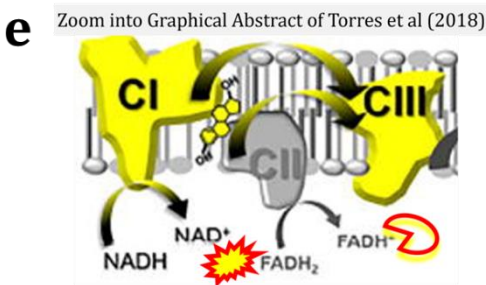
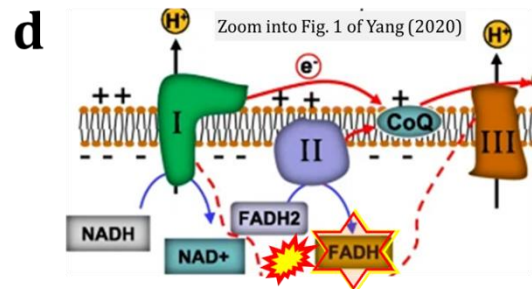
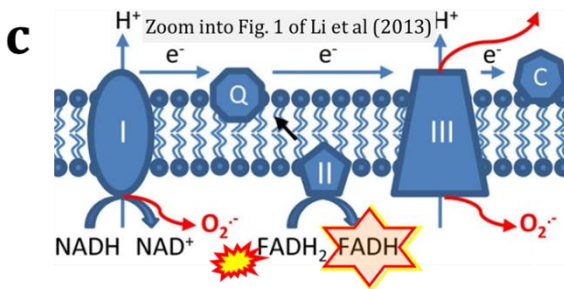
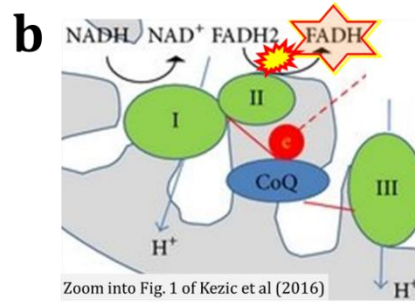
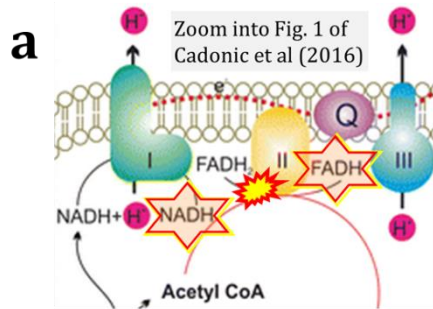
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Supplement 6

FADH₂ or FADH as substrate of CII and FADH, FADH⁺, or FAD⁺ as product (Figure S6)

- a-d  FADH₂ →  FADH (4)
- e  FADH₂ →  FADH⁺ (5)
- f  FADH → (6)
- g  FADH → FAD⁺ (7)
- h  FADH → FAD⁺ + H⁺ (8)
- i  FADH → FAD⁺ + 2H⁺ (9)



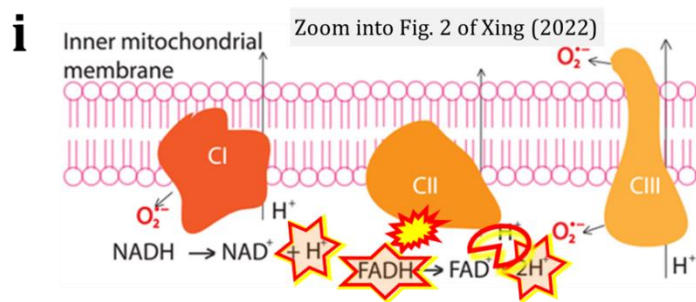


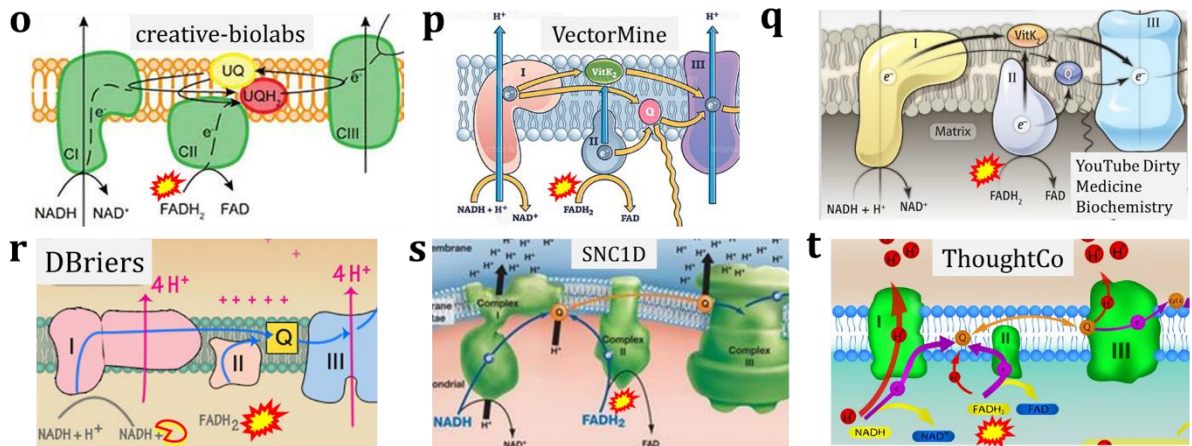
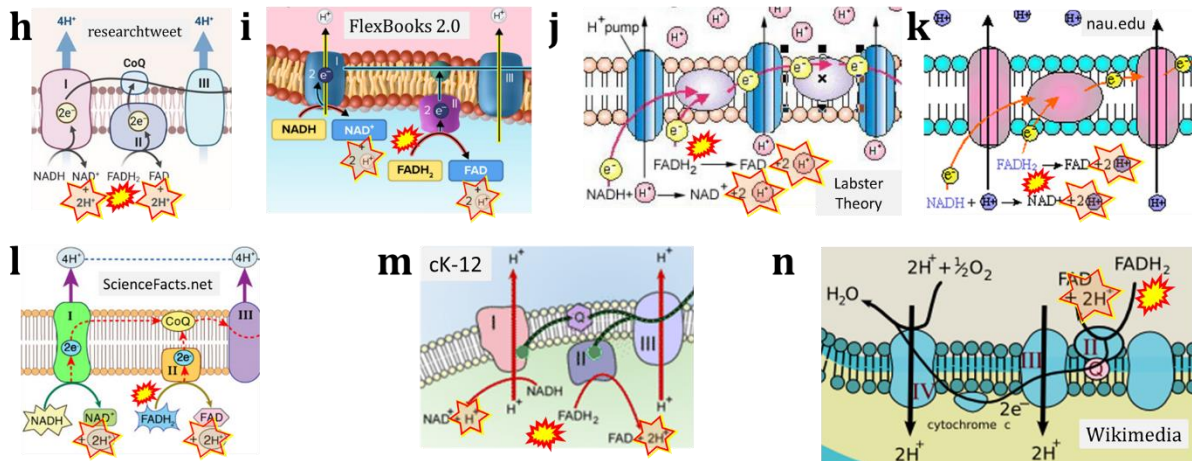
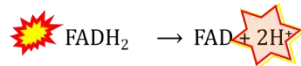
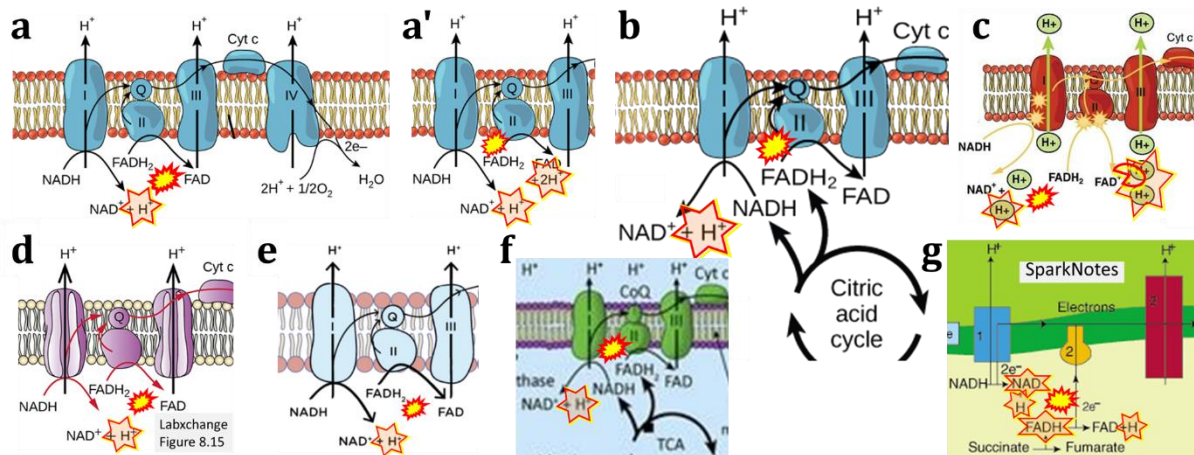
Figure S6. Complex II ambiguities: FADH₂ as substrate of CII and FADH or FADH⁺ as product. Sequence of publications from 2001 to 2023 according to (4) to (9). See References for Figure S6.

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Supplement 7

FADH₂ or FADH as substrate of CII in websites (Figure S7)



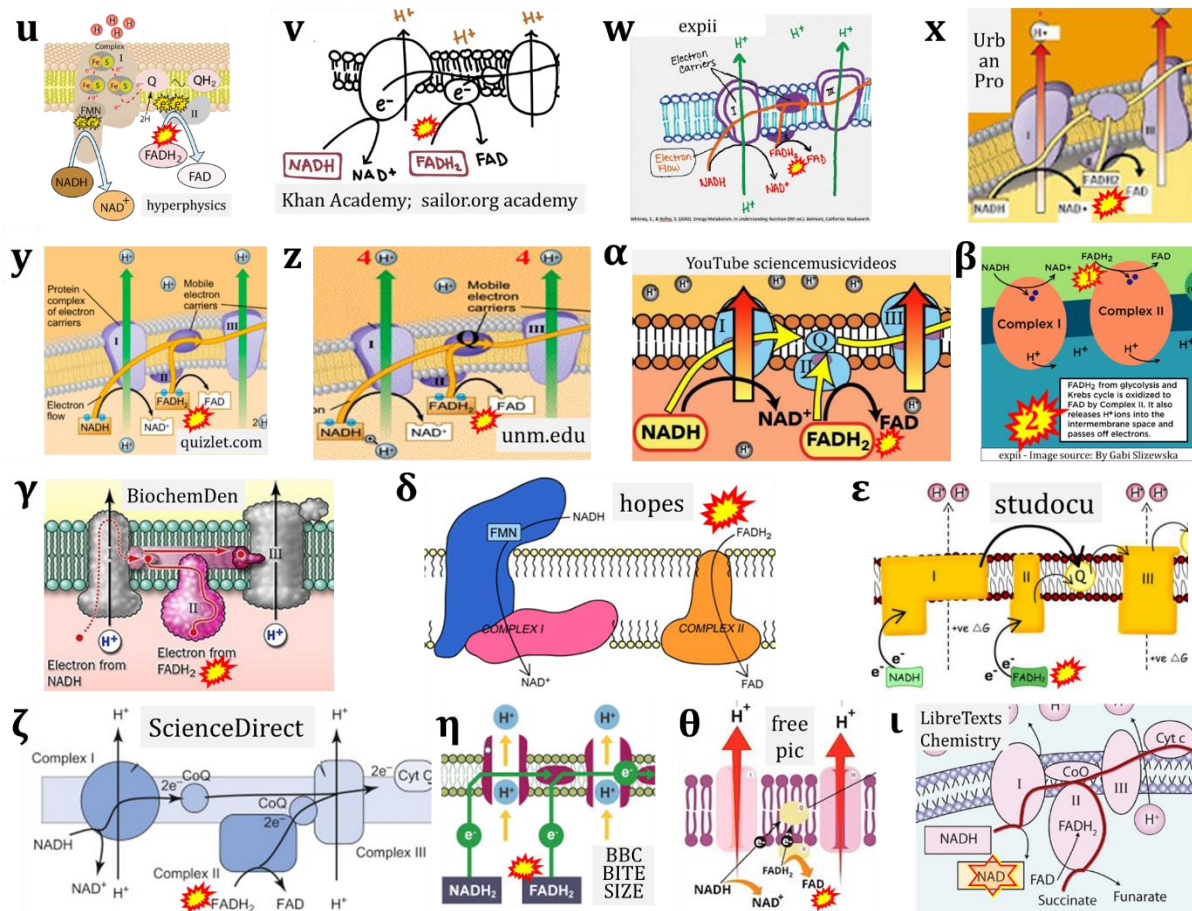


Figure S7. Complex II ambiguities in graphical representations on FADH₂ as a substrate of Complex II in the canonical forward electron transfer. (a, b, d-f, o-θ) FADH₂ → FAD; (a', h-n) FADH₂ → FAD+2H⁺; (c) FADH₂ → FAD⁺ + 2H⁺; and (g) FADH → FAD+H; should be corrected to FAD → FADH₂. NADH → NAD⁺ is frequently written in graphs without showing the H⁺ on the left side of the arrow, except for (p-r). (a-f, m) NADH → NAD⁺+H⁺; (g) NADH → NAD+H; (h, i, l) NADH → NAD⁺+2H⁺; (j, k) NADH+H⁺ → NAD⁺+2H⁺; and (t) NADH → NAD should be corrected to NADH+H⁺ → NAD⁺ (Eq. 3a). Weblinks #: (a) 1-8; (a') 9-10; (b) 1-6,9-11; (c) 11-17; (d) 18; (e) 19; (f) 20; (g) 21; (h) 22-23; (i) 24; (j) 25; (k) 26; (l) 27; (m) 28; (n) 11,29; (o) 30; (p) 31-32; (q) 33; (r) 34; (s) 35; (t) 12,22,36; (u) 37; (v) 9,10; (w) 11; (x) 38; (y) 39; (z) 40; (α) 41; (β) 11; (γ) 42; (δ) 43; (ε) 44; (ζ) 45; (η) 46; (θ) 47; (l) 48.

Weblinks for Figure S7 (retrieved 2023-03-21 to 2023-05-04)

- 1 (a,b) <https://openstax.org/books/biology/pages/7-4-oxidative-phosphorylation> - OpenStax Biology (CC BY 3.0) - Fig. 7.10, Fig. 7.12.
- 2 (a,b) <https://opentextbc.ca/biology/chapter/4-3-citric-acid-cycle-and-oxidative-phosphorylation/> - Charles Molnar, Jane Gair, Concepts of Biology - 1st Canadian Edition, BCCampus - Fig. 4.19.
- 3 (a,b) <https://www.pharmaguideline.com/2022/01/electron-transport-chain.html> - Pharmaguideline
- 4 (a,b) <https://www.texasgateway.org/resource/74-oxidative-phosphorylation> - Texas Gateway - Fig. 7.11, Fig. 7.13.
- 5 (a,b) <https://opened.cuny.edu/courseware/lesson/639/overview> - CUNY
- 6 (a,b) <https://courses.lumenlearning.com/wm-biology1/chapter/reading-electron-transport-chain/> - lumen Biology for Majors I - Fig. 1, Fig. 3.

- 7 (a) [https://bio.libretexts.org/Bookshelves/Introductory and General Biology/Book%3AGeneral Biology \(Boundless\)/07%3ACellular Respiration/7.11%3AOxidative Phosphorylation - Electron Transport Chain](https://bio.libretexts.org/Bookshelves/Introductory_and_General_Biology/Book%3AGeneral_Biology_(Boundless)/07%3ACellular_Respiration/7.11%3AOxidative_Phosphorylation_-_Electron_Transport_Chain) - LibreTexts Biology – Fig. 7.11.1
- 8 (a) <https://brainbrooder.com/lesson/254/7-4-1-electron-transport-chain> - Brain Brooder
- 9 (a',b,v) <https://www.khanacademy.org/science/ap-biology/cellular-energetics/cellular-respiration-ap/a/oxidative-phosphorylation-etc> - Khan Academy - Image modified from "Oxidative phosphorylation: Fig. 1", by OpenStax College, Biology (CC BY 3.0) / Image modified from "Oxidative phosphorylation: Fig. 3," by Openstax College, Biology (CC BY 3.0)
- 10 (a',b,v) <https://learn.saylor.org/mod/page/view.php?id=32815> -Saylor Academy
- 11 (b,c,n,w,β) <https://www.exprii.com/t/electron-transport-chain-summary-diagrams-10139> - exprii - Image source: By CNX OpenStax / By OpenStax College CC BY 3.0, via Wikimedia Commons / Whitney, Rolfes 2002 / By User:Rozzychan CC BY-SA 2.5, via Wikimedia Commons
- 12 (c,t) <https://www.thoughtco.com/electron-transport-chain-and-energy-production-4136143> - ThoughtCo / extender01 / iStock / Getty Images Plus
- 13 (c) <https://commons.wikimedia.org/w/index.php?curid=30148497> - wikimedia 30148497 - Anatomy & Physiology, Connexions Web site. <http://cnx.org/content/col11496/1.6/>, 2013-06-19
- 14 (c) <https://biologydictionary.net/electron-transport-chain-and-oxidative-phosphorylation/> - biologydictionary.net 2018-08-21
- 15 (c) <https://www.quora.com/Why-does-FADH2-form-2-ATP> - Quora
- 16 (c) <https://teachmephysiology.com/biochemistry/atp-production/electron-transport-chain/> - TeachMePhysiology - Fig. 1. 2023-03-13
- 17 (c) <https://www.toppr.com/ask/question/short-long-answer-types-what-is-the-electron-transport-system-and-what-are-its-functions/> - toppr
- 18 (d) <https://www.labxchange.org/library/items/lb:LabXchange:005ad47f-7556-3887-b4a6-66e74198fbcf:html:1> - Labxchange - Figure 8.15 credit: modification of work by Klaus Hoffmeier
- 19 (e) <https://jackwestin.com/resources/mcat-content/oxidative-phosphorylation/electron-transfer-in-mitochondria> - Jack Westin MCAT Courses
- 20 (f) <https://videodelivery.net/79e91c40bf96f9692560fa378c5086b6/thumbnails/thumbnail.jpg> - videodelivery
- 21 (g) <https://www.sparknotes.com/biology/cellrespiration/oxidativephosphorylation/section2/> - SparkNotes
- 22 (h,t) <https://researchtweet.com/mitochondrial-electron-transport-chain-2/> - researchtweet
- 23 (h) <https://microbenotes.com/electron-transport-chain/> - Microbe Notes
- 24 (i) <https://flexbooks.ck12.org/cbook/ck-12-biology-flexbook-2.0/section/2.28/primary/lesson/electron-transport-bio/> - FlexBooks - CK-12 Biology for High School- 2.28 Electron Transport, Fig. 2
- 25 (j) https://theory.labster.com/Electron_Transport_Chain/ - Labster Theory
- 26 (k) <https://www2.nau.edu/~fpm/bio205/u4fg36.html> - nau.edu
- 27 (l) <https://www.sciencefacts.net/electron-transport-chain.html> - ScienceFacts
- 28 (m) <https://www.ck12.org/biology/electron-transport/lesson/The-Electron-Transport-Chain-Advanced-BIO-ADV/> - cK-12
- 29 (n) https://commons.wikimedia.org/wiki/File:Mitochondrial_electron_transport_chain.png - Wikimedia

[%3A The Citric Acid Cycle and Electron Transport](#) - LibreTexts Chemistry - The Citric Acid Cycle and Electron Transport – Fig. 12.4.3

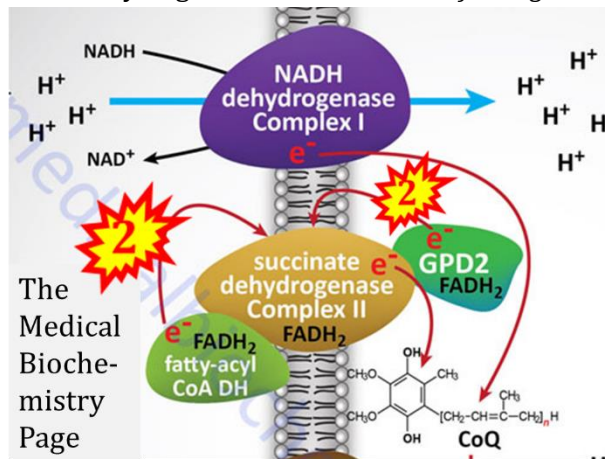
Supplement 8

Weblinks on FAO and CII (retrieved 2023-03-21 to 2023-05-02)

49 <https://conductscience.com/electron-transport-chain/> - Conduct Science: "In Complex II, the enzyme succinate dehydrogenase in the inner mitochondrial membrane reduce $FADH_2$ to FAD^+ . Simultaneously, succinate, an intermediate in the Krebs cycle, is oxidized to fumarate." - Comments: FAD does not have a positive charge. $FADH_2$ is the reduced form, it is not reduced. And again: In CII, FAD is reduced to $FADH_2$.

50 <https://themedicalbiochemistrypage.org/oxidative-phosphorylation-related-mitochondrial-functions/> - The Medical Biochemistry Page: 'In addition to transferring electrons from the $FADH_2$ generated by SDH, complex II also accepts electrons from the $FADH_2$ generated during fatty acid oxidation via the fatty acyl-CoA dehydrogenases and from mitochondrial glycerol-3-phosphate dehydrogenase (GPD2) of the glycerol phosphate shuttle' (Figure S8).

Figure S8. Fatty acid oxidation and glycerophosphate dehydrogenase erroneously shown to feed $FADH_2$ into Complex II. Weblink #50.



51 <https://www.chem.purdue.edu/courses/chm333/Spring%202013/Lectures/Spring%202013%20Lecture%2037%20-%2038.pdf> - CHM333 LECTURES 37 & 38: 4/27 – 29/13 SPRING 2013 Professor Christine Hrycyna - Acyl-CoA dehydrogenase is listed under 'Electron transfer in Complex II'.

Supplement 9

CII as a proton pump (Figure S9)

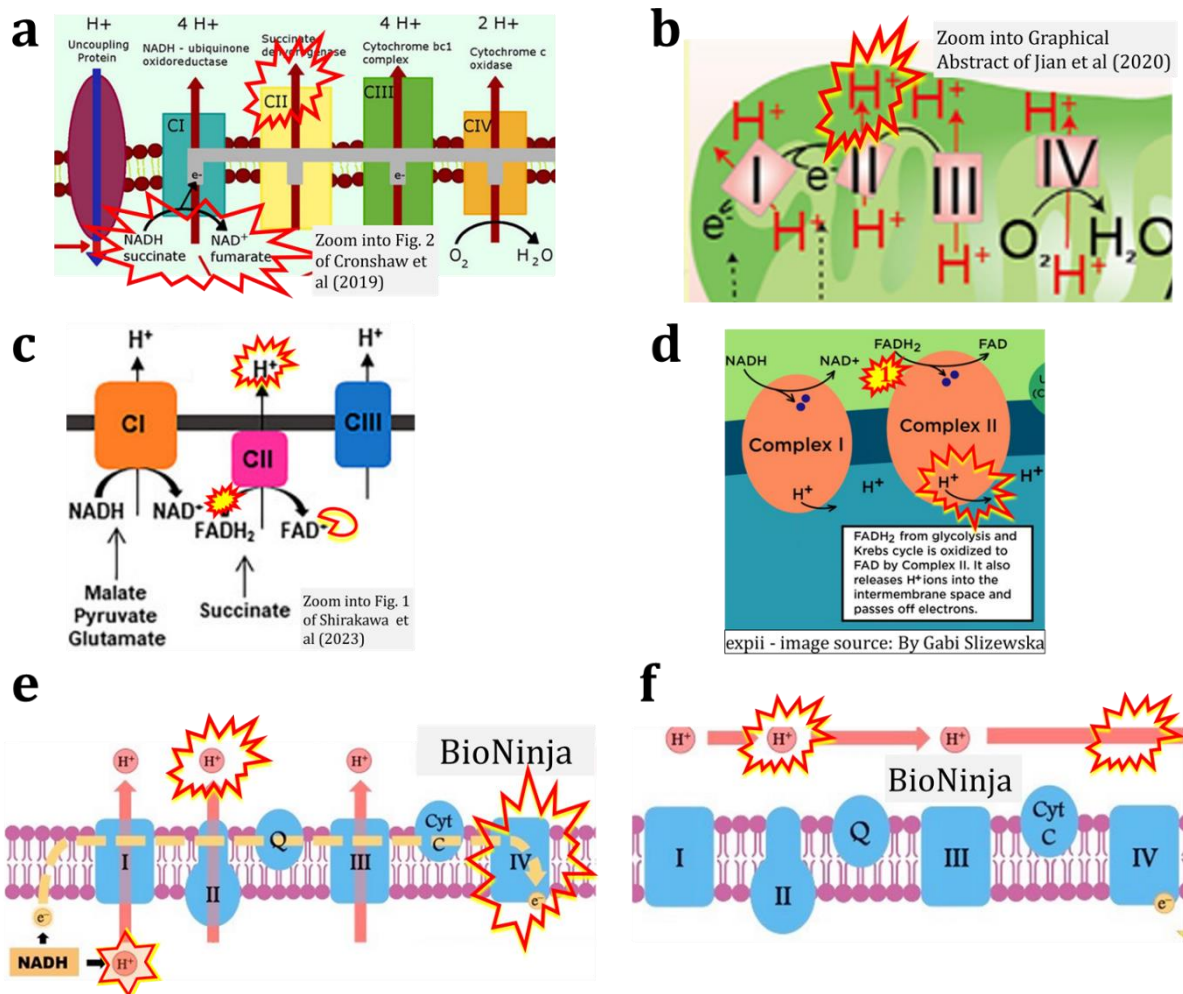


Figure S9. Complex II as a proton pump.

- a** Cronshaw M, Parker S, Arany P (2019) Feeling the heat: evolutionary and microbial basis for the analgesic mechanisms of photobiomodulation therapy. **Photobiomodul Photomed Laser Surg** 37:517-26. <https://doi.org/10.1089/photob.2019.4684>
- b** Jian C, Fu J, Cheng X, Shen LJ, Ji YX, Wang X, Pan S, Tian H, Tian S, Liao R, Song K, Wang HP, Zhang X, Wang Y, Huang Z, She ZG, Zhang XJ, Zhu L, Li H (2020) Low-dose sorafenib acts as a mitochondrial uncoupler and ameliorates nonalcoholic steatohepatitis. **Cell Metab** 31:892-908. <https://doi.org/10.1016/j.cmet.2020.04.011>
- c** Shirakawa R, Nakajima T, Yoshimura A, Kawahara Y, Orito C, Yamane M, Handa H, Takada S, Furihata T, Fukushima A, Ishimori N, Nakagawa M, Yokota I, Sabe H, Hashino S, Kinugawa S, Yokota T (2023) Enhanced mitochondrial oxidative metabolism in peripheral blood mononuclear cells is associated with fatty liver in obese young adults. **Sci Rep** 13:5203. <https://doi.org/10.1038/s41598-023-32549-w>
- d** <https://www.expil.com/t/electron-transport-chain-summary-diagrams-10139> - expii - Image source: By Gabi Slizewska: 'FADH₂ from glycolysis and Krebs cycle is oxidized to FAD by Complex II. It also releases H⁺ ions into the intermembrane space and passes off electrons' (retrieved 2023-05-04).
- e,f** <https://ib.bioninja.com.au/higher-level/topic-8-metabolism-cell/untitled/electron-transport-chain.html> - BioNinja (retrieved 2023-05-04).